

UNIVERSITY “ALEXANDRU IOAN CUZA” OF IAȘI  
THE FACULTY OF BIOLOGY  
DOCTORAL SCHOOL OF BIOLOGY

**Research on biological parameters of some species of the genus  
*Viola* L.: bioresources with complex potential for valorisation**

**PhD Thesis Summary**

**Scientific advisor:**

**UNIVERSITY PROFESSOR Ph.D. EMERITUS  
MARIA – MAGDALENA ZAMFIRACHE**

**Ph.D. Student:**

**ROSENHECH ELIDA**

**IAȘI  
2025**

# Summary

## Introduction

### Current state of research

1. History of research on *Viola* L. genus
2. The taxonomic variability of the studied species
  - 2.1. Subgenus *Viola* s.lat.
  - 2.2. Subgenus *Melanium* Ging.
3. Material, methods and working techniques

### Personal results

4. Morpho-anatomical research on some species of subgenus *Viola*
  - 4.1. Subsection *Viola* (= *Uncinatae* Kupffer, *Scapigeræ* W.Becker, *Curvato-pedunculatæ* W.Becker)
    - 4.1.1. Morphometric analyses of taxa of the *V. odorata* L., *V. suavis* Bieb., *V. alba* Bess.
    - 4.1.2. Anatomy analysis of taxa of the *V. odorata* L., *V. suavis* Bieb., *V. alba* Bess.
  - 4.2. Subsection *Rostratæ* Kupffer (W. Becker)
    - 4.2.1. Morphometric analyses of taxa of the *V. canina* L.
    - 4.2.2. Anatomy analysis of taxa of the *V. canina* L.
5. Morpho-anatomical research on some species of subgenus *Melanium*
  - 5.1. Morphometric analyses of taxa of the *V. declinata* W. et. K., *V. dacica* Borbás, *V. arvensis* Murray, *V. kitaibeliana* Shult. and *V. tricolor* L.
  - 5.2. Anatomy analyses of taxa of the *V. declinata* W. et. K., *V. dacica* Borbás, *V. arvensis* Murray, *V. kitaibeliana* Shult. and *V. tricolor* L.
6. Quantitative analyses of assimilatory pigments
7. Total quantitative analyses of phenols
8. Total quantitative analyses of flavonoid compounds
9. Total quantitative analyses of anthocyanin pigments
10. Determination of antioxidant capacity
11. Taxa of the genus *Viola* L., bioresources with complex potential for valorisation
12. Discussions
  - 12.1. Species discrimination based on morphometric indices
    - 12.1.1. Subgenus *Viola*
    - 12.1.2. Subgenus *Melanium*
  - 12.2. Species discrimination based on anatomical structure
    - 12.2.1. Subgenus *Viola*

12.2.2. Subgenus *Melanium*

12.3. The taxonomic confirmation of the species

12.4. The content of assimilatory pigments

12.5. Total phenols content

12.6. Total flavonoid content

12.7. Total anthocyanin pigments content

12.8. Assessment of the antioxidant potential

12.8. Assessment of the complex potential for the valorisation of *Viola*  
species

Conclusion

Appendix

References

## Introduction

Species of the genus *Viola* – violet, pansy – are spread across all continents, from temperate to tropical zones. Of the total of 550-600 species of this genus, 28 identified species grow spontaneously in Romania (Sârbu *et al.*, 2013; Toma and Ivănescu, 2013; Marcussen *et al.*, 2022).

The interest in using species of the genus *Viola* L. in the pharmaceutical industry stems from traditional medicine practices.

Internationally, studies on the biochemistry of the genus *Viola* have highlighted their pharmaceutical potential (Hammami *et al.*, 2011, 2012; Hellinger *et al.*, 2014; Saint-Lary *et al.*, 2014).

From an ecological point of view, these plants play an important role in maintaining biodiversity. (Knudsen *et al.*, 2006). Species of the subgenus *Melanium* are known to be metallophytes with a role in soil decontamination and ecosystem restoration. (Karatoteva *et al.*, 2014).

The main objective of this study is to supplement current scientific information with new data on the biology of some taxa belonging to the genus *Viola* existing in our spontaneous flora, collected from the North-Eastern Region of the country.

Three well-known and extensively studied species were examined (*V. odorata* L., *V. tricolor* L. and *V. arvensis* Murr.) in order to complete some data regarding their biology. These species are to be used as a benchmark in comparative research with other lesser-known species belonging to the Romanian plant heritage. *V. declinata* W. et K. and *V. dacica* Borb. have a special status, considered to be endemic to the Wooded Carpathians, Eastern and Southern Carpathian Mountains (Hurdu *et al.*, 2012; Alexiu, 2013).

Taxonomic complexity and large-scale intraspecific hybridization cause confusion regarding the systematic classification of species (Marcussen *et al.*, 2022), which is why this paper also aims to address some aspects regarding taxonomic variability through comparative research of some biological parameters. Nine species of the genus *Viola* L. were studied: *V. arvensis* Murr., *V. canina* L., *V. dacica* Borb., *V. declinata* W. et K., *V. kitaibeliana* Schult., *V. odorata* L., *V. suavis* Bieb. and *V. tricolor* L.

In this context, the present work aims to conduct a comparative study of some biological and biochemical parameters of the respective taxa, in order to complete the state of knowledge regarding elements of the variability of the genus *Viola* L.

Preliminary research on the variability of the genus *Viola* L. led to the crystallization of the following main objectives that are the subject of the thesis:

- comparative research of some **morpho-anatomical parameters**, with emphasis on species that present phenotypic plasticity, in order to complete data from current scientific literature on some elements regarding their variability;
- research into **phytochemical parameters** with potential for pharmaceutical use, in order to complete data from current scientific literature;
- research on the **potential for valorisation** of species as bioresources.

## 1. History of research on *Viola* L. genus

The interest in using plants of this genus in the pharmaceutical industry stems from traditional medicine practices. Currently, several species are recognized as exhibiting antimicrobial and antifungal, antiplasmodial and anthelmintic activity (Hammami *et al.*, 2011, 2012). Also, different species have antihypertensive, antidyslipidemic, anticancer, analgesic, antipyretic, anti-inflammatory, diuretic, anti-HIV and antiasthmatic activity. (Muhammad *et al.*, 2012).

In Romania, research on *Viola* species has highlighted the presence of volatile oils in *V. tricolor* and *V. arvensis* (Toiu *et al.*, 2006), salicylic acid content in *V. tricolor*, *V. arvensis* and *V. declinata* (Toiu *et al.*, 2008), phenolic compounds content in *V. declinata*, (Toiu *et al.*, 2009), flavonoids content in *V. arvensis* and *V. declinata* (Toiu *et al.*, 2017), as well as analyses of methanolic extracts for saponins, mucilage and carotenoid pigments (Toiu *et al.*, 2009).

The latest studies and technologies use bioactive compounds specific to the *Viola* genus in modern treatments against neurodegenerative diseases and cancer (Pränting *et al.*, 2010; Chandra *et al.*, 2015; Moliner *et al.*, 2019; Dayani *et al.*, 2022). Cyclotides and flavonoid compounds isolated from *Viola sp.* extracts are used as antibacterial active principles (Pränting *et al.*, 2010; Zarrabi *et al.*, 2013). Flavonoids, such as violantine, are integrated into fabrics through modern encapsulation technologies, for patients who need clothing with ultraviolet protection or treatments for skin conditions (e.g., to treat eczema) (Zimniewska, 2019).

## 2. The taxonomic variability of the studied species

Genus *Viola* L. (*Violaceae* Batsch.) is divided into 16 subgenera (Sârbu *et al.*, 2013; Marcussen *et al.*, 2015). Plants of this genus grow in almost all ecological zones, except Antarctica, but mainly in the temperate zones of both hemispheres. The taxonomic epicentre is in the Mediterranean area, East Asia and North America. (Marcussen *et al.*, 2022).

The species studied fall into:

- subgenus *Viola* s.lat.;
  - section *Viola* - *V. odorata* L., *V. suavis* Bieb. and *V. alba* Bess.
  - section *Rostratae* - *V. canina* L.
- subgenus *Melanium* Ging., section *Bracteolatae* – *V. declinata* W. et. K., *V. dacica* Borbás, *V. arvensis* Murray, *V. kitaibeliana* Shult. and *V. tricolor* L.

## 2.1. Subgenus *Viola* s.lat.

### ■ *Viola odorata* L.

The plant is annual (flowers in March-April), but can also be biannual, spreading throughout the country. It is characterized by a dense articulated rhizome, long lateral stolons, leaves arranged in a rosette, ovate-lanceolate stipules (Ciocîrlan, 1988; Ballard et al., 1999; Jonsell et al., 2009; Chifu et al., 2001, 2006). Due to the diverse habitat conditions, the species has a great morphological variability. The color of the petals is generally purple, but sometimes it can be lilac, white, pale pink, sulphur, blue, reddish, variegated (Ciocîrlan, 1988; Jonsell et al., 2009).



Figure. 1. *V. odorata* L.  
(Bârnova, Iași, Romania, 2020,  
original foto)

### ■ *Viola suavis* Bieb.

It is a perennial plant (blooms in March-April) spread from the plains (steppe zone) to the mountain area (beech zone), and prefers nutrient-rich soils (Ciocîrlan, 1988; Marcussen and Nordal, 1998; Jonsell et al., 2009).

In general, the species is characterized by short, densely jointed rhizomes, 4-5 mm thick, aboveground stolons 15 cm long, approximately 2 mm thick, sometimes underground stolons, long fimbriated stipules and bracteoles inserted on the lower half of the flowering peduncle (Mereďa Jr et al., 2008; Jonsell et al., 2009; Sârbu et al., 2013). Due to the different types of habitats, the taxon has high phenotypic plasticity, just like *V. odorata* L. and *V. alba* Bess.



Figure. 2. *V. suavis* Bieb.  
(Bârnova, Iași, Romania,  
2019, original foto)

■ ***Viola alba* Bess.**

It is a perennial plant (blooms in March-April), widespread up to 700 m altitude. It is common in the steppe zone and up to the beech forest area (Beldie *et al.*, 1955; Ciocârlan, 1988; Jonsell *et al.*, 2009).

It is characterized by flowers with white petals, almost ovate and acute leaves, linear-lanceolate and long-fimbriated stipules, light

green in colour, and fairly long stolons (30 cm) that do not root at the nodes. The rhizome is densely articulated (Jonsell *et al.*, 2009; Sârbu *et al.*, 2013).



Figure. 3. *V. alba* Bess.  
(Bârnova, Iași, Romania,  
2020, original foto)

■ ***Viola canina* L.**

It is part of the subsection *Rostratae* Kupffer (W. Becker).

The plants are perennial (flowering in May-June). It is common in the area of oak forests up to the spruce layer (1200 – 1700 m altitude). It prefers sunny habitats with moderate shade (Chifu *et al.*, 2001, 2006; Jonsell *et al.*, 2009).

The species is characterized by its herbaceous stem, free and long stipules, reaching up to the middle of the petiole. The style has a cylindrical shape and the stigma is rostrate.

Due to the high variety of habitats inhabited, plants may have stipules and leaves of different sizes and general shapes, or flowers of various colours (Beldie *et al.*, 1955; Jonsell *et al.*, 2009; Espeut, 2020).



Figure. 4. *V. canina* L.  
(Bârnova, Iași, Romania,  
2020, original foto)



## 2.2 Subgenus *Melanium* Ging.

### ■ *Viola declinata* W. et. K.

This plant is perennial and blooms in July-August. The species is characterized by the elliptic-lanceolate middle leaves and the linear-lanceolate upper ones. The stipules are finely sectioned up to the midrib (Sârbu *et al.*, 2013).

It is common in the spruce to juniper layer, in meadows (Tomescu and

Harasemciuc, 2018). It is characteristic for pastures with *Nardus stricta*. It grows in juniper areas, forest clearings, sunny places, on rocks and scree. It is also found on secondary meadows established after the clearing of spruce trees (Chifu *et al.*, 2006; Togor and Burescu, 2013).

This species is endemic to the Wooded Carpathians, Eastern, and Southern Carpathians (Ciocârlan, 1988; Coldea *et al.*, 2009; Hurdu *et al.*, 2012).

In Romania, as a result of personal observations carried out in the field between 2015-2022, we can report the presence of this species in alpine and subalpine meadows from different subdivisions of the Carpathian Mountains, such as: Călimani Mountains, trails to 12 Apostoli and Pietrosul Călimanilor Peak; Rodnei Mountains at Gârgălau Saddle, Știol plateau, Iezerul Pietrosului and in the vicinity of Craiu Peak; Bucegi Mountains at Țigănești Saddle and Scara Peak plateau; Piatra Craiului Mountains, trail to Măgura Mare Peak.



Figure. 5. *V. declinata* W. et. K.  
(Pietrosul Călimanilor Peak,  
Romania, 2020, original foto)

### ■ *Viola dacica* Borbás

Perennial plant that blooms in July-August. It is common in the spruce and juniper areas. It is endemic to Europe (Coldea *et al.*, 2009; Hurdu *et al.*, 2012).

The species is characterized by its middle leaves being broad-ovate and the upper ones being oblong-ovate. The stipules are less divided, 1/2-1/3 of their width, the undivided part being wider than the length of the segments. (Sârbu *et al.*, 2013).

Personal observations from 2015-2022, in Romania, highlight the presence of this species in meadows and pastures (alpine and subalpine) in: Călimani Mountains, from Neagra Șarului to 12 Apostoli, and Pietrosul Călimanilor Peak, Rodnei Mountains on the peaks of Lucurel and Craia, Sângeorz Hillock, Obârșia Rebrii Saddle; Giumalău Mountains, Ceahlău Mountains, Făgăraș Mountains in the areas of Fereastră Mare, Urlea Peak, Dara Peak and Hârtopul Darei Peak.



Figure. 6. *V. dacica* Borbás (Sângeorz Hillock, Rodnei Mountains, Romania, 2020, original foto)

### ■ *Viola arvensis* Murray

It is cosmopolitan, annual, blooms in the months of May-August, the plant having a distribution throughout Romania. The species is frequent in the steppe area up to the beech zone, widespread in the hill area, to the limit of cereal crops and up to the subalpine zone (Beldie *et al.*, 1955; Ciocârlan, 1988; Sârbu *et al.*, 2013).

Characteristic for this species is the length of the corolla equal to or shorter than the calyx, with flowers of 1-1.5 cm long. The middle leaves are elongated and ovate-elliptic (Sârbu *et al.*, 2013).

The species is morphologically similar to *V.*



Figure. 7. *V. arvensis* Murray (Cioatele, Vaslui, Romania, 2018, original foto)

*tricolor* L. and *V. kitaibeliana* Shult. which is why identification errors may occur (Jonsell *et al.*, 2009; Espeut, 2020).

#### ■ *Viola kitaibeliana* Shult.

It is annual and blooms in the months of April-July. Generally, we find it on rocky or sandy coasts, meadows and bushes, it prefers arid places. It is sporadic in the steppe area and up to the *Quercus* zone (up to 600-700 m altitude) (Beldie *et al.*, 1955; Magrini and Scoppola, 2015).

Characteristic of the species are the flower length of 0.4-0.8 cm and broad-ovate middle leaves (Sârbu *et al.*, 2013).

Phenotypic plasticity causes species identification problems because it has morphological characteristics similar to *V. arvensis* Murray and *V. tricolor* L. (Espeut, 2020).

This species is considered endangered in Italy, which is why it has a special status (Magrini and Zucconi, 2020).



Figure. 8. *V. kitaibeliana* Shult. (Târgu Neamț, Neamț, Romania, 2019, original foto)

#### ■ *Viola tricolor* L.

It is a cosmopolitan species, the plant is spread throughout the entire territory of Romania, and almost throughout the entire European territory. It blooms in the months of May-August. It is annual, sometimes biannual. This species is widespread in the hilly area (200-300 m minimum altitude), up to the subalpine level (Chifu *et al.*, 2006; Tomescu and Harasemciuc, 2018; Espeut, 2020).

The species is characterized by the length of the corolla which is longer than the



Figure. 9. *V. tricolor* L. (Fălticeni, Suceava, Romania, 2019, original foto)

calyx and the pinnate-lobed stipules with the terminal segment larger than the lateral ones (Sârbu *et al.*, 2013).

Due to habitats with different ecological requirements, some morphological characteristics show phenotypic varieties influenced by environmental conditions, mainly by the soil qualities (Jonsell *et al.*, 2009; Słomka *et al.*, 2008, 2011; Bezlova *et al.*, 2012).

### **3. Material, methods and working techniques**

#### **1. The plant material investigated**

The investigated species were collected from different spontaneous flora habitats located in four counties from Romania: Iași, Neamț Suceava and Vaslui.

The plant material belonging to the selected species was harvested during the flowering phenophase, from three different populations, over a period of two consecutive years.

The plant material was determined taxonomically according to Sârbu *et al.*, 2013 and verified by Biologist Ph.D. Irinia Irina, de la Herbarium of the Faculty of Biology, University „Alexandru Ioan Cuza” of Iași, where vouchers of the taxa taken into study are also deposited.

#### **2. Techniques for morphological analyses**

The morphometry was performed on herborized specimens. Measurements were performed according to the methods described in the specialized literature (Marcussen and Nordal, 1988; Marcussen *et al.*, 2001; Marcussen, 2003; Hodálová *et al.*, 2008).

#### **3. Methods and techniques for plant histoanatomy**

The plant material, preserved in 70% ethyl alcohol, was sectioned with a hand microtome and a botanical razor. The samples thus obtained were studied and photographed with an optical microscope (Niță *et al.*, 1997; Toma, 2003; Cutler and Botha 2007).

#### 4. Determination of water and dry matter content

The classic method of removing water by repeatedly drying the plant material was used (Boldor *et al.*, 1982; Hodgson *et al.*, 2011).

#### 5. Quantitative analyses of assimilatory pigments

The assimilatory pigments were extracted using the Mayer–Bertenrath method (Boldor *et al.*, 1982).

#### 6. Quantitative analyses of anthocyanin pigments

For the quantitative determination of anthocyanin pigments, the pH differential method was used (Fuleki and Francis, 1968).

#### 7. Plant extracts

The dried plant material containing the aerial part of the plants, without flowers, was mechanically ground. The solvents used to obtain extracts were distilled water and 50% ethanol. The plant extracts were made for four concentrations: 0.5%, 1%, 2.5% and 5% w/v. (dry material weight/volume).

#### 8. Quantitative analyses of phenols

For the quantitative evaluation of the phenol content, we used the reagent Folin-Ciocalteu method (Herald *et al.*, 2012).

#### 9. Quantitative analyses of flavonoids

To evaluate the total amount of flavonoids, we used the classic analysis method taken from Jia *et al.*, 1999, and Herald *et al.*, 2012.

#### 10. Determination of antioxidant capacity

Estimation of antioxidant activity was performed using the stable free radical 2,2-diphenyl-1-picrylhydrazyl, based on a colorimetric reaction (Blois, 1985; Molyneux, 2003; Siatka and Kašparová, 2010).

#### 11. Statistical data processing

Statistical data processing was carried out using Microsoft Excel 2015, XLSTAT and OriginLab Pro.

## 12. PCR analyses

PCR analysis was performed within the Molecular Biology and Metagenomics Laboratory, University „Ștefan cel Mare” of Suceava.

### *a) Genomic DNA extraction*

Genomic DNA was extracted from dried plant material using the Wizard Genomic DNA Purification Kit (Promega).

### *b) Genomic region selection and primer design*

Specific primers for each species were selected based on sequences available in genetic databases and specialized literature (Ballard *et al.*, 1998; Hildebrandt *et al.*, 2006; Malécot *et al.*, 2007; Mered'a *et al.*, 2011; Pomon *et al.*, 2016; Tamura *et al.*, 2021).

### *c) PCR Protocol*

It was used GoTaq Green Master Mix (Promega) on the equipment Applied Biosystems SimpliAmp Thermal Cycler.

#### 4. Morpho-anatomical research on some species of subgenus *Viola*

##### 4.1. Subsection *Viola* (= *Uncinatae* Kupffer, *Scapigeræ* W.Becker, *Curvato-pedunculatae* W.Becker)

##### 4.1.1. Morphometric analyses of taxa of *V. odorata* L., *V. suavis* Bieb., *V. alba* Bess.

Table 4.1. Morphometric indices analyzed for *V. odorata* L., *V. suavis* Bieb., *V. alba* Bess. (mm, average values  $\pm$  standard deviation)

Morphometry	<i>V.odorata</i>	<i>V. suavis</i>	<i>V.alba</i>
length (L) petiole	26.54 $\pm$ 0.97	36.6 $\pm$ 0.32	27.33 $\pm$ 0.42
length (L) stipule	10 $\pm$ 0	10.73 $\pm$ 0.16	11 $\pm$ 0.67
width (l) stipule	2.53 $\pm$ 0.17	3.13 $\pm$ 0.37	1 $\pm$ 0
ratio L/l stipule	4.11 $\pm$ 0.2	3.62 $\pm$ 0.3	11.00 $\pm$ 0.5
leaf area (A)	956.43 $\pm$ 26.74	684.67 $\pm$ 61.93	488.33 $\pm$ 22.93
length (L) leaf	59.29 $\pm$ 2.53	60.63 $\pm$ 2.68	49.04 $\pm$ 4.24
width (l) leaf	34.59 $\pm$ 0.97	26.99 $\pm$ 1.05	24.46 $\pm$ 1.69
leaf perimeter (P)	210.44 $\pm$ 3.57	166.37 $\pm$ 9.13	147.85 $\pm$ 2.26
ratio L/l leaf	1.70 $\pm$ 0.1	2.38 $\pm$ 0.2	2.13 $\pm$ 0.3
length (L) peduncle	61.07 $\pm$ 0.82	32.47 $\pm$ 0.39	53.4 $\pm$ 1.91
length (L) bracts	3.87 $\pm$ 0.2	4.73 $\pm$ 0.5	3.47 $\pm$ 0.1
width (l) bracts	0.65 $\pm$ 0.1	1.00 $\pm$ 0	0.50 $\pm$ 0.0
ratio L/l bracts	6.53 $\pm$ 0.6	4.73 $\pm$ 0.5	6.93 $\pm$ 0.3
length (L) flower	12.6 $\pm$ 0.51	10.84 $\pm$ 0.1	15 $\pm$ 0
length (L) sepal	5.13 $\pm$ 0.13	6 $\pm$ 0	4.2 $\pm$ 0.08
width (l) sepal	2.52 $\pm$ 0.08	2 $\pm$ 0	1 $\pm$ 0
ratio L/l sepal	2.08 $\pm$ 0.1	3.00 $\pm$ 0.0	4.20 $\pm$ 0.1

#### **4.1.2. Anatomy analyses of taxa of *V. odorata* L., *V. suavis* Bieb., *V. alba* Bess.**

From an anatomical point of view, the differences between these three species were as follows:

- *root*: thickness of the xylem, presence or absence of xylem parenchyma cells, presence or absence of periderm; only in *V. odorata* the structure remains primary, with visible absorbent hairs at the rhizoderm level;
- *rhizome*: the degree of thickening of the xylem vessels, respectively, the presence and frequency of cellulosic parenchyma xylem cells;
- *scapus type stem*: presence (*V. alba*), respectively absence in the bark of air cavities; number of conductive fascicles: 4 at *V. odorata*, 2 at *V. suavis*, 2 at *V. alba*;
- *leaf blade*: bicellular tector bristles are present at the level of the leaf blade veins, with the terminal cell very long at *V. odorata*, stomata protrude above the external level of the epidermis at *V. odorata* and *V. suavis*, at *V. alba* the cuticle is thick.

#### **4.2. Subsection *Rostratae* Kupffer (W. Becker)**

##### **4.2.1. Morphometric analyses of taxa of *V. canina* L**

A number of 10 morphometric indices associated with the leaf (petiole, stipule and leaf blade) and aerial stems were analysed. A number of 8 morphometric indices were considered for the flower.

##### **4.2.2. Anatomy analysis of taxa of *Viola canina* L.**

- *root*: the structure, typically secondary, is the result of the activity of both lateral meristems: the cambium and the phellogen;
- *rhizome*: the structure is of secondary origin, but with a thin periderm and a very thick central cylinder. The pith, extremely thin, parenchymatic-cellulosic, is penetrated by short rows of primary xylem;
- *aerial stem*: the outline is semicircular, with two slightly prominent adaxial ridges and a flat adaxial face. The central cylinder is thick, comprising 7-8 conductive fascicles of different sizes;
- *leaf*:



The petiole - the outline is approximately elliptical modified by two strongly divergent latero-adaxial ridges.

Lamina – amphistomatic, the midrib protrudes visibly on both sides and the lateral ones of order I, only on the lower side of the leaf blade .

## 5. Morpho-anatomical research on some species of subgenus *Melanium*

### 5.1. Morphometric analyses of taxa of *V. declinata* W. et. K., *V. dacica* Borbás, *V. arvensis* Murray, *V. kitaibeliana* Shult. and *V. tricolor* L.

Table 5.2. Morphometric indices analyzed for *V. declinata* W. et. K., *V. dacica* Borbás, *V. arvensis* Murray, *V. kitaibeliana* Shult. and *V. tricolor* L. (mm, average values  $\pm$  standard deviation)

Morphometry	<i>V. dacica</i>	<i>V. declinata</i>	<i>V. arvensis</i>	<i>V. kitaibeliana</i>	<i>V. tricolor</i>
length (L) petiole	3.60 $\pm$ 0.2	2.40 $\pm$ 0.2	5.20 $\pm$ 0.4	5.07 $\pm$ 0.6	8.33 $\pm$ 0.9
length (L) stipule	5.27 $\pm$ 0.1	4.80 $\pm$ 0.1	8.27 $\pm$ 0.4	9.07 $\pm$ 0.3	14.93 $\pm$ 1.0
width (l) of terminal lobe of the stipule	1.08 $\pm$ 0.0	1.00 $\pm$ 0.0	1.47 $\pm$ 0.1	1.53 $\pm$ 0.1	2.70 $\pm$ 0.3
leaf area (A)	149.45 $\pm$ 12.7	90.20 $\pm$ 4.2	178.93 $\pm$ 12.4	149.53 $\pm$ 12.2	180.07 $\pm$ 14.2
length (L) leaf	29.11 $\pm$ 1.1	22.62 $\pm$ 1.7	30.13 $\pm$ 1.6	31.83 $\pm$ 1.0	28.77 $\pm$ 1.5
width (l) leaf	10.24 $\pm$ 0.6	7.16 $\pm$ 0.5	10.31 $\pm$ 0.5	9.89 $\pm$ 0.6	11.46 $\pm$ 0.3
leaf perimeter (P)	58.14 $\pm$ 4.3	47.71 $\pm$ 2.0	69.08 $\pm$ 3.2	57.95 $\pm$ 1.5	66.06 $\pm$ 3.1
Ratio L/l lamina	3 $\pm$ 0.2	3.72 $\pm$ 0.6	3.07 $\pm$ 0.3	3.31 $\pm$ 0.1	2.50 $\pm$ 0.1

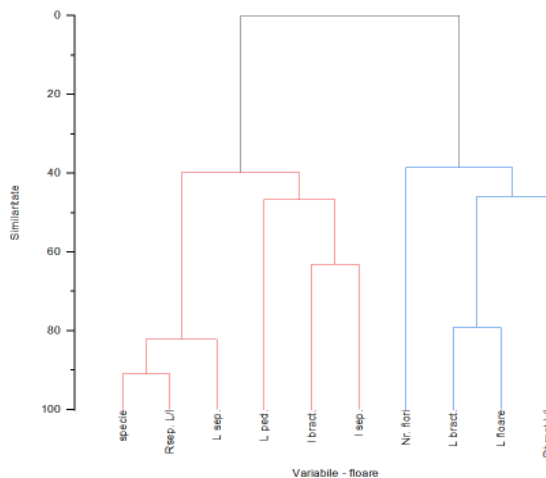


Figure 5. 1. Multivariate cluster analysis for flower morphometric indices for *V. declinata* W. et. K., *V. dacica* Borbás, *V. arvensis* Murray, *V. kitaibeliana* Shult. and *V. tricolor* L.

## 5.2. Morphometric analyses of taxa of the *V. declinata* W. et. K., *V. dacica* Borbás, *V. arvensis* Murray, *V. kitaibeliana* Shult. and *V. tricolor* L.

From an anatomical point of view, the main distinguishing features between these are the following:

- *root*: elliptical outline at *V. tricolor*, circular to other species;
- *rhizome*: is present only at *V. dacica* and *V. declinata*;
- *stem*: number of conductive fascicles - 10 for *V. dacica*, 7 for *V. declinata*, 10 for *V. tricolor*, 8 for *V. arvensis*, 8 or 9 for *V. kitaibeliana*;
- *petiole*: *V. tricolor* has 7 conductive fascicles centrally located and 3 in each adaxial wing; *V. arvensis* and *V. kitaibeliana* have 3 conductive fascicles: one central and one in each wing; *V. dacica* has 7 centrally located and *V. declinata*, 3 conductive fascicles centrally located.

## 6. Quantitative analyses of assimilatory pigments

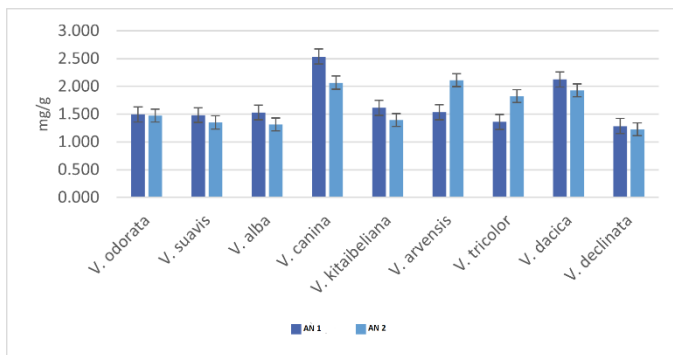


Figure 6.2. Total quantitative content of assimilatory pigments (chlorophyll a + chlorophyll b + carotenoid pigments) for the two consecutive years (mg/g), in the analysed species.

## 7. Total quantitative analyses of phenols

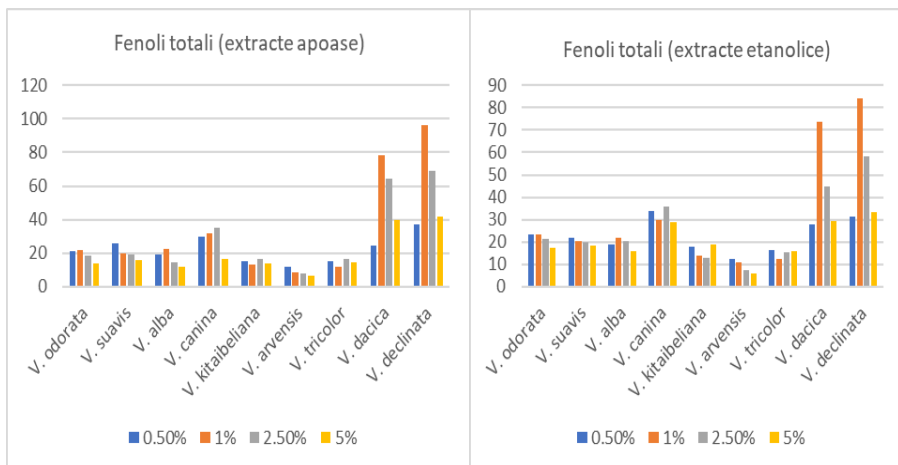


Figure 7.9. The total amount of polyphenols determined for aqueous and ethanolic extracts for concentrations of 0.5%, 1%, 2.5%, respectively 5%, for nine species of the genus *Viola* L. (mg/g GAE/dry wt.)

## 8. Total quantitative analyses of flavonoid compounds

Two types of extracts were analysed, aqueous and ethanolic (50%), with four concentrations being made for each: 0,5% (w/v), 1% (w/v), 2,5% (w/v) and 5% (w/v). The highest values were recorded for *V. canina* and *V. kitaibeliana*.

Comparing the two types of extracts, it is observed that the ethanolic extracts have a higher extraction efficiency of total flavonoid compounds. The highest values were recorded for the alcoholic extracts made for *V. canina*:  $101.12 \pm 2.40$  mg/g QE/dry wt. (0,5%),  $87.17 \pm 0.73$  mg/g QE/dry wt. (1%),  $97.61 \pm 0.31$  mg/g QE/dry wt. (2,5%) and  $80.78 \pm 0.62$  mg/g QE/dry wt. (5%).

The lowest values, for all concentrations in both types of extracts, were observed mainly in the *V. arvensis*.

## 9. Total quantitative analyses of anthocyanin pigments

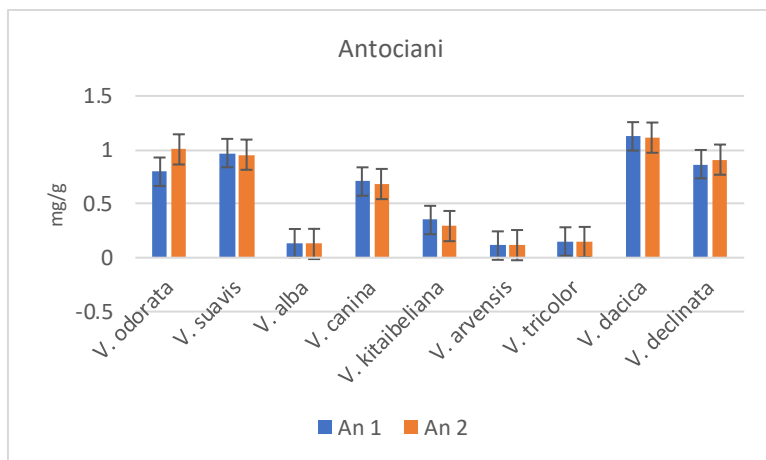


Figure 9.3. Comparative graph of the average values obtained for anthocyanin compounds following analyses carried out for nine species of the genus *Viola* L., in two consecutive years (mg/g dry wt.)

## **10. Determination of antioxidant capacity**

Testing the antioxidant capacity of plant extracts revealed significant activity.

Low values of the inhibition rate were recorded in species harvested from agricultural areas, *V. arvensis* (~17%-24% for aqueous extracts and ~18%-25% for the ethanolic extracts), and *V. tricolor* (~19-22% for aqueous extracts and ~19%-29% for the ethanolic extracts).

The highest values of the inhibition rate were ~30%-47% for aqueous extracts and ~46%-54% for the ethanolic extracts (*V. dacica*) and ~35%-63% for aqueous extracts and ~44%-55% for the ethanolic extracts (*V. declinata*).

## **11. Taxa of the genus *Viola* L., bioresources with complex potential for valorisation**

### **11.1. Bioresources of valuable compounds in the pharmaceutical and cosmetic industry**

The total content of polyphenols recorded by analysing the experimentally prepared aqueous extracts presented the highest values in species collected from mountain areas. (*V. declinata*, *V. dacica*), with values between 78-84 mg/g GAE/dry wt. (1% w/v). The lowest values recorded for aqueous extracts were 8-13 mg/g GAE/dry wt. (1%), for *V. arvensis* and *V. kitaibeliana*. Comparatively, the quantitative analysis of total polyphenols for alcoholic extracts revealed that the highest values were 94-95 mg/g GAE/dry wt. (1%) (*V. declinata*, *V. dacica*) and the lowest values were 10-19 mg/g GAE/dry wt. (1%) (*V. arvensis*, *V. kitaibeliana*).

The class of polyphenolic compounds is recognized as having bactericidal, antifungal, anti-inflammatory and, very importantly, antioxidant properties.

Based on the results obtained, an overall analysis can be undertaken on the potential for valorisation of certain species of *Viola*.

## **11.2. Bioresources with ecological impact**

This genus contains taxa that develops in very different biocenoses, from cosmopolitan species with diverse growth area, to endemic species with strict requirements or adapted to difficult environmental conditions. The adaptability of some taxa, such as *V. odorata* and *V. suavis*, allows them to develop in semi-natural habitats or habitats modified by human activity, gardens or parks (Ciocîrlan, 1988; Chifu *et al.*, 2001, 2006). Resistance to polluted soils and especially the accumulation of these pollutants in the plant body, gives species of the subgenus *Melanium* a major role in ecosystem restoration activities. (Hermann *et al.*, 2013).

## **12. Discussions**

### **12.1. Species discrimination based on morphometric indices**

The morphological analyses of the investigated species revealed common characters between the species, but also discriminatory elements between them. Our observations are, for the most part, in accordance with the specialized literature. The differences recorded are due to the fact that the species under study present a high phenotypic variability.

### **12.2. Species discrimination based on anatomical structure**

The general anatomical structure is consistent with the description made for the genus *Viola*, in specialized literature. The observed anatomical differences may be useful in supplementing the specialized literature for identifying taxa by microscopic means.

### **12.3. The taxonomic confirmation of the species**

Confirming that the taxa analysed were correctly identified, a molecular analysis of nuclear ribosomal DNA was performed.

#### 12.4. The content of assimilatory pigments

The content of assimilatory pigments, as well as the ratio between chlorophylls a and b, are influenced by the light incidence on the leaf surface, which is why the highest values were recorded in species collected from alpine and subalpine meadows (*V. canina* and *V. dacica*).

#### 12.5. Total phenols content

The total content of polyphenols recorded, analysing the experimentally prepared alcoholic and aqueous extracts, was highest in species collected from mountain areas *V. declinata* and *V. dacica*, closely followed by *V. canina*. Species collected from agricultural areas and semi-natural pastures presented the lowest values (*V. arvensis*, *V. tricolor* and *V. kitaibeliana*).

In general, the experimentally obtained data is similar to those presented in the specialized literature.

#### 12.6. Total flavonoid content

The quantitative analysis of total flavonoids from aqueous and alcoholic extracts prepared from the tested *Viola* plants revealed that the maximum values were recorded for *V. canina*, and the minimum values for *V. arvensis*.

Analysing the similarity of the results obtained with those in the specialized literature, the values obtained by us partially correlate with the published data.

#### 12.7. Total anthocyanin pigments content

The highest value obtained were in species with intensely purple petals (*V. dacica*, *V. declinata*, *V. canina*) and blue violet (*V. odorata*, *V. suavis*). The results obtained are partially correlated with published data, the fluctuation of quantitative values of anthocyanin compounds both between species and between populations of the same species may be due to harvesting from different habitats, soil quality or water stress.

### 12.8. Assessment of the antioxidant potential

Species of the genus *Viola* contain a significant number of compounds from the polyphenol class, which is why the extracts produced have a high antioxidant capacity. The inhibition rate values, for aqueous and ethanolic extracts, were high for mountain species, and lower for species harvested from agricultural areas (*V. kitaibeliana*, *V. tricolor* and *V. arvensis*).

### 13.9. Assessment of the complex potential for the valorisation of *Viola* species

Polyphenolic compounds are recognized in modern medicine for their anti-inflammatory and antibacterial effects, as well as their strong antioxidant capacity, qualities that have led to their use as adjuvants in the treatments of multiple conditions.

From an ecological point of view, the species have significant importance, both as cosmopolitan plants, growing on various substrates, including anthropized soils, and as taxa that occupy habitats with difficult and polluted environmental conditions.

## Conclusions

The main objective of the research carried out was to analyse the taxa belonging to a number of nine species of the genus *Viola*, subgenres *Viola* and *Melanium*. Starting from these species, the aim of the work was to enrich the specialized literature with new data on the biology of some taxa belonging to this genus, present in the spontaneous flora of the geographical area of the North-Eastern Region of Romania.

- The analysis of **phytochemical parameters** was based on the premise that species of the genus *Viola* are appreciated in traditional medicine at an international level, which is why aqueous and ethanolic extracts were made with four concentrations (0.5%, 1%, 2.5% and 5%) using dried, mechanically ground plant material. Due to the applied working methodology, the efficiency of the extraction of biochemical compounds was



limited by the type of solvent and the chosen concentration. The difference between the values obtained practically and those reported in the specialized literature may be due either to the difference in environmental factors in the habitat areas, to the harvesting period, or to the combination of these two premises.

- Comparative analyses of some **morphometric** and **anatomical parameters** have highlighted new elements that can be taken into account for species discrimination, the data obtained having a pronounced novelty character.
- **The phytochemical parameters** analysed were the *content of polyphenols, flavonoids, anthocyanin pigments and determination of antioxidant capacity*. The results obtained identified significant quantities of these classes of compounds in the analysed plant material. For the total amount of phenols and flavonoids, low values were determined in species collected from agricultural areas: *V. arvensis*, *V. tricolor* and *V. kitaibeliana*, and high values in species harvested from mountainous areas: *V. declinata*, *V. dacica*, *V. canina*. Quantitative analysis of anthocyanin pigments revealed low values in species with white, cream and pale-yellow flowers: *V. alba* and *V. arvensis*, and high values were identified in species with purple flowers: *V. declinata*, *V. dacica*, *V. canina*, *V. odorata* and *V. suavis*. The evaluation of antioxidant potential highlighted the antioxidant capacity of both aqueous and ethanolic plant extracts, with the latter recording higher values compared to the aqueous ones. The results obtained identified in the analysed plant material significant quantities of these compounds with **potential for pharmaceutical use**.
- Personal results, as well as those presented in the specialized literature, highlight the complex **potential for valorisation** of species of the genus *Viola*.
- From an **ecological** point of view, species of the genus *Viola* occupy a diverse number of habitats, including niche ones, such as alpine screes, to anthropized habitats, reality that contributes directly and/or indirectly to the health of ecosystems, soil quality, increasing the quality of the vegetation cover and the production of biomass available for insects and

herbivores. The resistance to polluted soils and especially the accumulation of heavy metals in the vegetative organs give species of the genus *Viola* a major role in soil decontamination and ecosystem restoration. All these attributes prove the special importance of these species, as well as the need for their conservation.

- **Morphometric** research on endemic species (*V. declinata* and *V. dacica*) and **anatomy** (*V. suavis*, *V. canina*), along with the identification of morphometric indices applicable for discriminating the analysed species, as well as research on the chemical composition of taxa belonging to the genus *Viola* collected from the North-East Region of Romania have a pronounced **novelty character**; these data enrich the national specialized literature with valuable information for completing the taxonomic determination keys of the studied taxa, for phytochemists and phytopharmacists, as well as for specialists working in the direction of environmental protection, engaged in the effort of its conservation and biological restoration.

The results obtained can be used as premises for new **research directions** in order to deepen knowledge regarding the biology of the *Viola* genus and to obtain a more detailed picture of how its taxa interact with the habitats they inhabit. Biochemical testing of extracts obtained from the analysed plant material reveals the importance of developing more complex experimental models to streamline the extraction and isolation of biologically active compounds of interest. Also, expanding the analyses to investigate a larger number of biocompounds may provide information necessary for new therapeutic applications of the analysed taxa. The evaluation of new species of the genus *Viola*, as well as their hybrids, can contribute to the identification of new bioresources with complex potential for valorisation and can highlight relevant information regarding the conservation status of the Romanian plant gene pool.

## **Acknowledgment**

I honour the memory of Mr. Academician Constantin TOMA (1935-2020), and I express my gratitude for the support and guidance provided in the research activity that formed the basis for the completion of this doctoral thesis.

## Appendix – selection



a.



b.



c.

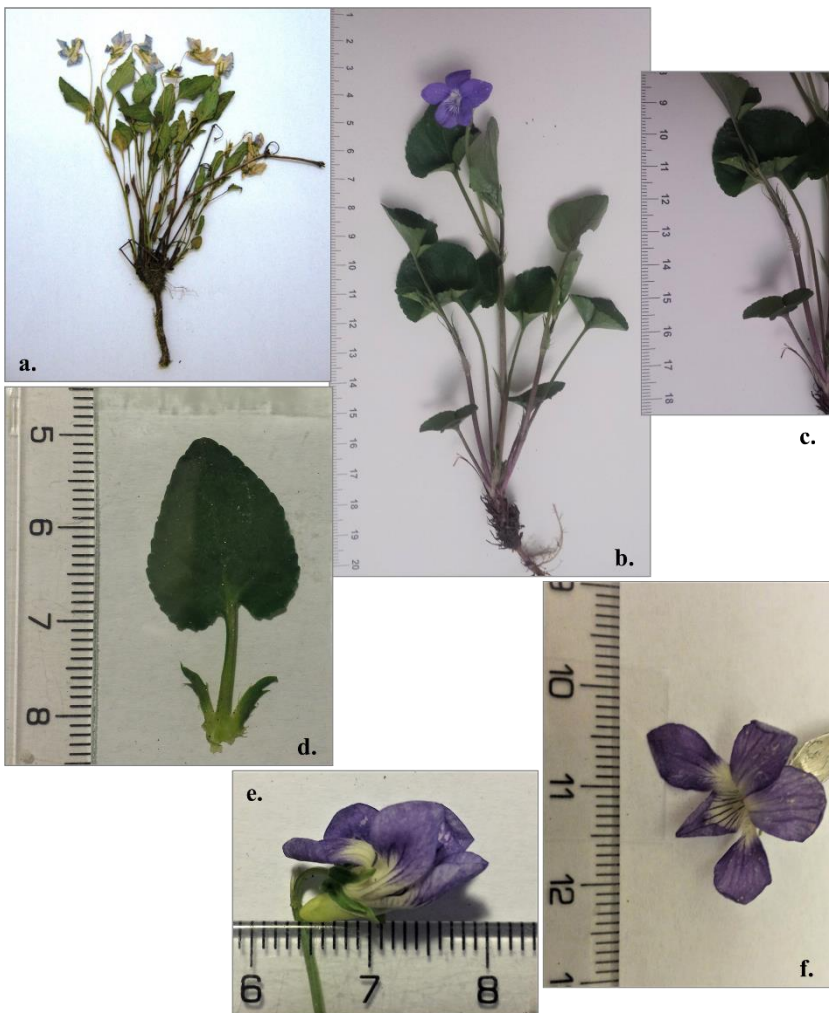


d.



e.

**Exhibit no. 7.** *V. canina* L.: - a./c. Giumalău Mountain-deciduous Forest edge; b./d. Giumalău Mountain - short grass meadow; 29.04.2021 - e. Rarău Mountain – coniferous forest, 30.04.2021 (original photo)



**Exhibit no. 8.** *V. canina* L. - a. general image, plant with multiple aerial stems, herborized; - b. general image; - c. detail showing the stipules; - d. leaf from the upper part of the aerial stem, with stipules; - e./f. flower (original photo)

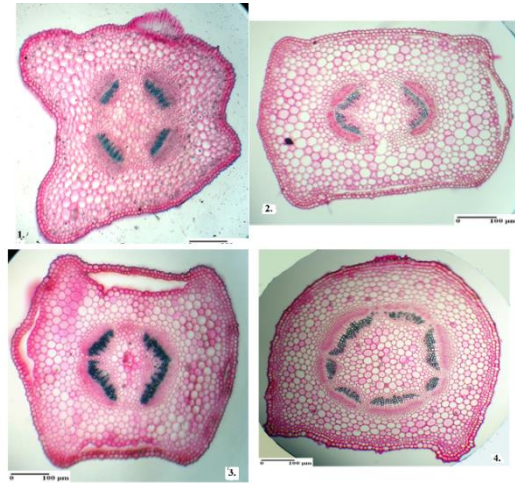


Figure. 10. Cross section through flower peduncle: 1. *V. odorata* L., 2. *V. suavis* Bieb., 3. *V. alba* Bess./ 4. Cross section through the stem *V. canina* L.

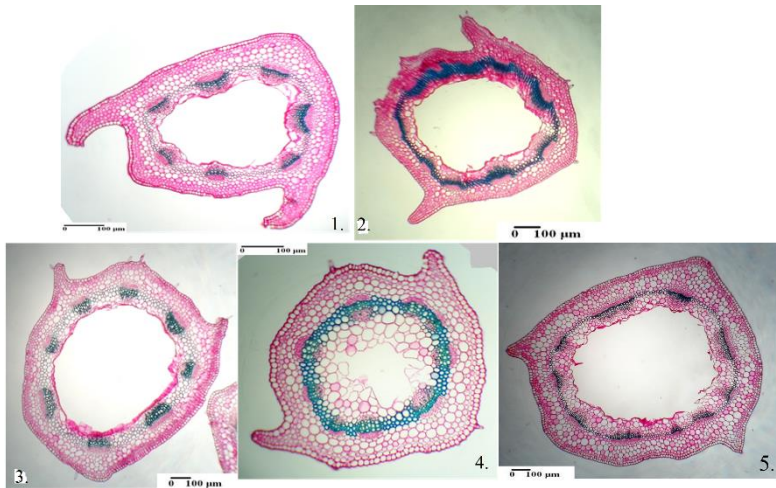


Figure. 11. Cross section through stem: 1. *V. declinata* Waldst. et Kit; 2. *V. dacica* Borbás; 3. *V. arvensis* Murr.; 4. *V. kitaibeliana* Schult.; 5. *V. tricolor* L.

## Appendix 2

### List of species of the genus *Viola* L. in Romania

The presence of a number of 28 species belonging to the genus *Viola* L. was reported on the territory of Romania (Sârbu *et al.*, 2013):

1. *V. alba* Bess. [2 ssp] – white pansies
2. *V. alpina* Jacq.
3. *V. ambigua* W. et K.
4. *V. arvensis* Murray (include *V. banatica* Kit. Ex Roem. et Schult.)
5. *V. biflora* L. – yellow pansies
6. *V. canina* L. [3 ssp]
7. *V. collina* Bess.
8. *V. dacica* Borb. – bird's claw
9. *V. declinata* W. et K. – bird's claw
10. *V. elatior* Fr.
11. *V. epipsila* Ledeb.
12. *V. hymettia* Boiss. et Heldr.
13. *V. hirta* L.
14. *V. joói* Janka.
15. *V. jordanii* Hanry
16. *V. kitaibeliana* Schult.
17. *V. mirabilis* L. – pansies
18. *V. odorata* L. - pansies
19. *V. palustris* L. [1 ssp]
20. *V. persicifolia* Schreb. (= *V. stagnina* Kit.; *V. lactea* Auct.)
21. *V. pumila* Chaix
22. *V. reichenbachiana* Jord. Ex Boreau (= *V. sylvestris* Lam.)
23. *V. riviniana* Rchb
24. *V. rupestris* F. W. Schm.
25. *V. sieheana* W. Beckr
26. *V. suavis* Bieb. (= *V. cyanea* Čelak, *V. pontica* W. Becker, *V. ignobilis* Grinț.) – pansies
27. *V. tricolor* L. [2 subsp.] – three spotted brothers
28. *V. uliginósa* Bess. (presence is not certain, reconfirmation required)

## References- selection

1. Ardelean, M., Cachiță-Cosma, D., Ardelean, A., Lădașiu, F. C., Lobiuc, A., Zamfirache, M. M., & **Rosenhech, E.** (2017). Cytological aspects and anthocyanin accumulation observed in *Sedum telephium* ssp. *maximum* L. callus." 13125-13134.
2. Ballard H. E., Paula Souza J., Wahlert G. A. (2014). Violaceae. Pp. 303-322 In: Kubitzki, K. (ed.), The families and genera of vascular plants. Springer-Verlag, Berlin
3. Ballard H. E., Paula-Souza J., Seidel R. (2015 ["2014"]). Violaceae. Pp. 1353-1356 In: Jørgensen, P. M., M. Nee, S. Beck (eds.), Catálogo de las Plantas Vasculares de Bolivia, Monographs in Systematic Botany. Missouri Botanical Garden, St. Louis, MO.
4. Ballard Jr, H. E., Sytsma, K. J., & Kowal, R. R. (1999). Shrinking the violets: phylogenetic relationships of infrageneric groups in *Viola* (Violaceae) based on internal transcribed spacer DNA sequences. *Systematic Botany*, 439-458.
5. Ballard, H. E. (2015). Violaceae. Pp. 593-611 In: Hammel, B., M. Grayum, C. Herrera & N. Zamora (eds.), Manual de Plantas de Costa Rica. Missouri Botanical Garden, St. Louis, MO.
6. Ballard, H. E., Jr., Sytsma, K. J., & Kowal, R. R. (1998). *Viola arvensis* internal transcribed spacer 1, complete sequence (Accession No. AF097242). GenBank, National Center for Biotechnology Information.
7. Ballard, H. E., Jr., Sytsma, K. J., & Kowal, R. R. (1998). *Viola odorata* internal transcribed spacer 2, complete sequence (Accession No. AF097296). GenBank, National Center for Biotechnology Information.
8. Beldie Al. *Viola*. In: Savulescu T. editor. (1955). Flora Republicii Populare Române, Vol III, Editura Academiei Republicii Populare Române, p. 553-625
9. Bezlova, D., Tsvetkova, E., Karatoteva, D., & Malinova, L. (2012). Content of heavy metals and arsenic in medicinal plants from recreational areas in Bulgarka Nature Park. *Genetics and Plant Physiology*, 2(1/2), 64-72.
10. Blois M.S. (1958). Antioxidant determinations by the use of a stable free radical. *Nature* 181, 1199 - 1200
11. Bojinescu, R. I., & Burescu, P. (2018). Phytocoenology and ecology of the *Nardus stricta* meadows edified by *Nardus stricta* and *Viola*



- declinata in Semenic Mountains. *Analele Universității din Oradea, Fascicula: Protecția Mediului*, 30, 107-116.
12. Boldor O., Raianu O., Trifu M. (1982). *Fiziologie vegetală. Lucrări practice*, Edit. Didactică și pedagogică, București: 5-8, 160-161
  13. Burescu, Ioan-Nuțu. L. (2015). Rare, endangered, vulnerable, endemic, relict plants and animals encompassing high conservation values for the forests of Vlădeasa mountains-the northern Apuseni mountains. *Analele Universității din Oradea, Fascicula Protecția Mediului* 25: 309-318
  14. Burnea, I., Popescu, I., Neamțu, G., Stancu, E., Lazăr, Ș. (1977), *Chimie și biochimie vegetală*, Edit. Didactică și Pedagogică, București, p.325-328
  15. Burzo Ioan. (1999). *Fiziologie vegetală: curs universitar*. Ed. Universității de Științe Agronomice și Medicină Veterinară, București
  16. Burzo, I. (2015). Radicalii liberi, rolul acestora și importanța substanțelor antioxidante din plante în menținerea sănătății umane. Ed. Elisaveros, București, p. 64-94, 109-110, 115-119
  17. Burzo, I., Simion, T., Crăciun, C., Voican, V., Dobrescu, A., Delian, E. (1999). *Fiziologia plantelor de cultură. Vol. I, Întreprinderea Editorial Poligrafică Știința, Chișinău*
  18. Butură, V. (1979). *Enciclopedie de etnobotanică românească*. Editura Științifică și Enciclopedică. Editura Științifică și Enciclopedică, București
  19. Cao, D. L., Zhang, X. J., Xie, S. Q., Fan, S. J., & Qu, X. J. (2022). Application of chloroplast genome in the identification of Traditional Chinese Medicine *Viola philippica*. *BMC genomics*, 23(1), 1-19.
  20. Castro-López, C., Ventura-Sobrevilla, J. M., González-Hernández, M. D., Rojas, R., Ascacio-Valdés, J. A., Aguilar, C. N., & Martínez-Ávila, G. C. (2017). Impact of extraction techniques on antioxidant capacities and phytochemical composition of polyphenol-rich extracts. *Food Chemistry*, 237, 1139-1148.
  21. Cesco, S., Mimmo, T., Tonon, G., Tomasi, N., Pinton, R., Terzano, R., Neumann, G., Weisskopf, L., Renella, G., Landi, L. and Nannipieri, P. (2012). Plant-borne flavonoids released into the rhizosphere: impact on soil bioactivities related to plant nutrition. A review. *Biology and Fertility of Soils*, 48, 123-149.

22. Cesco, S., Neumann, G., Tomasi, N., Pinton, R., & Weisskopf, L. (2010). Release of plant-borne flavonoids into the rhizosphere and their role in plant nutrition. *Plant and Soil*, 329, 1-25.
23. Çetlin, M. (2016). Changes in the amount of chlorophyll in some plants of landscape studies. *Kastamonu Üniversitesi Orman Fakültesi Dergisi*, 16(1), 239-245.
24. Chandra, D., Kohli, G., Prasad, K., Bisht, G., Punetha, V.D., Khetwal, K.S., Devrani, M.K., Pandey, H.K. (2015). Phytochemical and ethnomedicinal uses of family *Violaceae*. *Curr. Res. Chem*, 7, 44-52.
25. Chifu, T., Zamfirescu, O., Mânzu, C., & Şurubaru, B. (2001). *Botanică sistematică–Cormobionta*, Ed. Universităţii, „Alexandru Ioan Cuza”, Iaşi, 388-389.
26. Chifu, T.; Mânzu, Ciprian; Zamfirescu, Oana. (2006). *Flora şi vegetaţia Moldovei (României)*. Editura Universităţii “Alexandru Ioan Cuza”, Iaşi: 290-291
27. Chiriţoiu, M. (2007). Corologia plantelor rare şi vulnerabile din Carpaţii meridionali (2). In *Realizări şi perspective în horticultură, viticultură, vinificaţie şi silvicultură*” (Vol. 15, pp. 101-103).
28. Ciocîrlan, Vasile. (1988). *Flora ilustrată a României*, Vol. 1 *Determinarea şi descrierea speciilor spontane şi cultivate*. Editura Ceres: 9-58, 495-503
29. Coldea, G., Stoica, I. A., Puşcaş, M., Ursu, T., Oprea, A., & IntraBioDiv Consortium. (2009). Alpine–subalpine species richness of the Romanian Carpathians and the current conservation status of rare species. *Biodiversity and Conservation*, 18, 1441-1458.
30. Coombs E. R. (2003). *Violets: The history & cultivation of scented Violets*. BT. Batsford Publisher, a II-a ediţie, ISBN: 071348831 X (prima ediţie – 1981, Croom Helm Ltd.)
31. Creţu, R., Macovei, A., Băra, I. C., Ghiorgă, G., Verdeş, R., Iocob, E., Ionescu E. (2011). Physiological effects induced by the hydroalcoholic extract of *Viola tricoloris* herba (wild pansy aerial parts) on *Triticum aestivum* L., *Analele Ştiinţifice ale Universităţii „Alexandru Ioan Cuza”, Secţiunea Genetică şi Biologie Moleculară*, Tom XII: 125-132
32. Dakora, F. D. (1995). Plant flavonoids: biological molecules for useful exploitation. *Functional Plant Biology*, 22(1), 87-99.
33. Dalrymple, R. L., Kemp, D. J., Flores-Moreno, H., Laffan, S. W., White, T. E., Hemmings, F. A., & Moles, A. T. (2020).

- Macroecological patterns in flower colour are shaped by both biotic and abiotic factors. *New Phytologist*, 228(6), 1972-1985.
34. Dani, R. S., & Shrestha, K. K. (2004). Two new records of *Viola* L.(Violaceae) for Nepal. *Himalayan Journal of Sciences*, 2(3), 48-50.
  35. Danu, M. A., Chifu, T. (2008). Contributions to the study of the class molinio-arhenatheretea r. tx. 1937 in the upper basin of River Dorna (Suceava County) (i); *Analele științifice ale Universității "Al. I. Cuza" Iași*, s. II a. Biologie vegetală, Tomul LIV, fasc. 1: 136-145
  36. Dastagir, G., Bibi, S., Uza, N. U., Bussmann, R. W., & Ahmad, I. (2023). Microscopic evaluation, ethnobotanical and phytochemical profiling of a traditional drug *Viola odorata* L. from Pakistan. *Ethnobotany Research and Applications*, 25, 1-24.
  37. Dayani, L., Varshosaz, J., Aliomrani, M., Dinani, M. S., Hashempour, H., & Taheri, A. (2022). Morphological studies of self-assembled cyclotides extracted from *Viola odorata* as novel versatile platforms in biomedical applications. *Biomaterials Science*, 10(18), 5172-5186.
  38. Doniță, N., Paucă-Comănescu, M., Popescu, A., Mihăilescu, S., & Biriș, I. A. (2005). *Habitatele din România*. București: Editura Tehnică Silvică.
  39. Duțu, M., Ardelean, A., Ardelean, M., Cachiță-Cosma, D., Marian, B., Lobiuc, A., & **Rosenhech, E.** (2016). Increasing the antioxidant activity, total phenolic and assimilatory pigments content by optimizing the in vitro growth conditions of *Lycium barbarum* plant. *Scientific Bulletin. Series F. Biotechnologies*, Vol. XX, 2016, 44-50
  40. Eckstein, R. L., Hölzel, N., & Danihelka, J. (2006). Biological Flora of Central Europe: *Viola elatior*, *V. pumila* and *V. stagnina*. *Perspectives in Plant Ecology, Evolution and Systematics*, 8(1), 45-66.
  41. Espeut, M. (2020). Revision of the genus *Viola* L.(Violaceae) in the Russian Far East with notes on adjacent territories. *Botanica Pacifica*, 9(1), 3-52.
  42. Farzad, M., Griesbach, R., Hammond, J., Weiss, M. R., & Elmendorf, H. G. (2003). Differential expression of three key anthocyanin biosynthetic genes in a color-changing flower, *Viola cornuta* cv. Yesterday, Today and Tomorrow. *Plant Science*, 165(6), 1333-1342.
  43. Fuleki T., Francis F. J. (1968). Quantitative methods for anthocyanins. *Journal of food science*, 33: 72-77

44. Gonçalves, A. F. K., Friedrich, R. B., Boligon, A. A., Piana, M., Beck, R. C. R., & Athayde, M. L. (2012). Anti-oxidant capacity, total phenolic contents and HPLC determination of rutin in *Viola tricolor* (L) flowers. *Free Radicals and Antioxidants*, 2(4), 32-37.
45. Hammami, I., Kamoun, N., & Rebai, A. (2011). Biocontrol of *Botrytis cinerea* with essential oil and methanol extract of *Viola odorata* L. flowers. *Arch. Appl. Sci. Res*, 3(5), 44-51.
46. Hashimoto, H., Uragami, C., & Cogdell, R. J. (2016). Carotenoids and photosynthesis. In *Carotenoids in nature: biosynthesis, regulation and function*, Stange, C., Ed.; Springer, Germany, 111-139.
47. Hayden, W. J., & Clough, J. (1990). Methyl salicylate secretory cells in roots of *Viola arvensis* and *V. rafinesquii* (Violaceae). *Castanea*, 65-70.
48. Hellinger, R., Koehbach, J., Fedchuk, H., Sauer, B., Huber, R., Gruber, C. W., & Gründemann, C. (2014). Immunosuppressive activity of an aqueous *Viola tricolor* herbal extract. *Journal of ethnopharmacology*, 151(1), 299-306.
49. Herald, T. J., Gadgil, P., & Tilley, M. (2012). High-throughput micro plate assays for screening flavonoid content and DPPH-scavenging activity in sorghum bran and flour. *Journal of the Science of Food and Agriculture*, 92(11), 2326-2331.
50. Hermann, B., Katarina, V. M., Paula, P., Matevž, L., Neva, S., Primož, P., Primož, V., Jeromel, L., & Marjana, R. (2013). Metallophyte status of violets of the section *Melanium*. *Chemosphere*, 93(9), 1844-1855.
51. Hildebrandt, U., Hoef Emden, K., Backhausen, S., Bothe, H., Bozek, M., Siuta, A., & Kuta, E. (2006). *Viola tricolor* subsp. *tricolor* clone a2 ITS1 5.8S ITS2 sequence (Accession No. DQ055406). GenBank, National Center for Biotechnology Information.
52. Hodálová, I., Mered'a Jr, P., Mártonfi, P., Mártonfiová, L., & Danihelka, J. (2008). Morphological characters useful for the delimitation of taxa within *Viola* subsect. *Viola* (Violaceae): a morphometric study from the West Carpathians. *Folia Geobotanica*, 43, 83-117.
53. Hodgson, J.G., Montserrat-Martí, G., Charles, M., Jones, G., Wilson, P., Shipley, B., Sharafi, M., Cerabolini, B.E.L., Cornelissen, J.H., Band, S.R. & Bogard, A. (2011). Is leaf dry matter content a better predictor of soil fertility than specific leaf area?. *Annals of botany*, 108(7), 1337-1345.

54. Hoyos Gomez. (2011). Towards an understanding of the basal evolution of Violaceae from an anatomical and morphological perspective. MSc thesis, University of Missouri – St. Louis, St. Louis MO
55. Hurdu, B. I., Pușcaș, M., Turtureanu, P. D., Niketić, M., Vonica, G., & Coldea, G. (2012). A critical evaluation of the Carpathian endemic plant taxa list from the Romanian Carpathians. *Contributii Botanice*, 47.
56. Ibraheem, R. M., Mhawesh, A. A., & Abood, K. W. (2018). Estimation of the whole flavonoid, antioxidant, antibacterial challenge concerning *Viola odorata* (banafsha) methanolic extract. *The Iraqi Journal of Agricultural Science*, 49(4), 655.
57. Jia, Z., Tang, M., & Wu, J. (1999). The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. *Food chem*, 64(4), 555-559.
58. Jităreanu, C. D. (2002). *Fiziologie vegetală*. Editura “Ion Ionescu de la Brad”, Iași
59. Jonsell, B., & Karlsson, T. (2010). Lectotypification of two Linnaean names for *Flora Nordica* Vol. 6. *Nordic Journal of Botany*, 28(2), 129-129.
60. Jonsell, B., Ericsson, S., Fröberg, L., Rostański, K., Snogerup, S., & Widén, B. (2009). Nomenclatural notes to *Flora Nordica* Vol. 6 (*Thymelaeaceae–Apiaceae*). *Nordic Journal of Botany*, 27(2), 138-140.
61. Jurca, T., Pallag, A., Marian, E., & Eugenia, M., (2019). The histo-anatomical investigation and the polyphenolic profile of antioxidant complex active ingredients from three *Viola* species. *Farm. J*, 67, 634-640.
62. Karatoteva, D., Tsvetkova, E., & Bezlova, D. (2014). Heavy metals and arsenic in soils from grasslands in Bulgarka Nature Park. *Bulgarian Journal of Agricultural Science*, 20(1), 73-78.
63. Karim, N., Khan, I., Abdelhalim, A., Khan, A., & Halim, S. A. (2018). Antidepressant potential of novel flavonoids derivatives from sweet violet (*Viola odorata* L): Pharmacological, biochemical and computational evidences for possible involvement of serotonergic mechanism. *Fitoterapia*, 128, 148-161.
64. Knudsen, J. T., Eriksson, R., Gershenzon, J., & Ståhl, B. (2006). Diversity and distribution of floral scent. *The botanical review*, 72(1), 1-120.

65. Koç, M. T., Dane, F., & Ekici, N. (2022). Comparative anatomical investigations on four *Viola* l.(Violaceae) taxa from European Turkey. In proceedings of IV. International agricultural, Biological & Life Science Conference Agbiol 2022, 322.
66. Li, Q., Wang, J., Sun, H. Y., & Shang, X. (2014). Flower color patterning in pansy (*Viola*× *wittrockiana* Gams.) is caused by the differential expression of three genes from the anthocyanin pathway in acyanic and cyanic flower areas. *Plant physiology and biochemistry*, 84, 134-141.
67. Linné Carl von, 1707-1778; translation by Freer Stephen - University of Oxford. (2003). *Linnaeus' Philosophia Botanica*. Oxford University Press Inc, New York: 78, 80-81, 85, 91, 108, 118, 120, 138, 161, 188-189, 252, 262, 287, 305.
68. Magrini, S., & Scoppola, A. (2015). Cytological status of *Viola kitaibeliana* (Section *Melanium*, Violaceae) in Europe. *Phytotaxa*, 238(3), 288-292.
69. Magrini, S., & Zucconi, L. (2020). Seed germination of the endangered *Viola kitaibeliana* and other Italian annual pansies (*Viola* Section *Melanium*, Violaceae). *Flora*, 30, 425.
70. Mahboubi, M., & Kashani, L. M. T. (2018). A Narrative study about the role of *Viola odorata* as traditional medicinal plant in management of respiratory problems. *Advances in Integrative Medicine*, 5(3), 112-118.
71. Malécot, V., Marcussen, T., Munzinger, J., Yockteng, R., & Henry, M. (2007). *Viola alba* subsp. *alba* voucher MH 043220 internal transcribed spacer 1, complete sequence (Accession No. DQ358849). GenBank, National Center for Biotechnology Information.
72. Marcussen T. (2003). Evolution, phylogeography and taxonomy within the *Viola alba* complex (Violaceae). *Plant Systematics and Evolution*, Austria, 237: 51-74
73. Marcussen T.; Nordal, I. (1998). *Viola suavis*, a new species in the Nordic flora, with analyses of its relation to other species in the subsection *Viola* (Violaceae). *Nordic Journal of Botany* 18: 221–237
74. Marcussen, T., & Borgen, L. (2000). Allozymic variation and relationships within *Viola* subsection *Viola* (Violaceae). *Plant Systematics and Evolution*, 223, 29-57.
75. Marcussen, T., Ballard, H. E., Danihelka, J., Flores, A. R., Nicola, M. V., & Watson, J. M. (2022). A revised phylogenetic classification for *Viola* (Violaceae). *Plants*, 11(17), 2224.

76. Marcussen, T., Borgen, L., & Nordal, I. (2005). New distributional and molecular information call into question the systematic position of the West Asian *Viola sintenisii* (Violaceae). *Botanical Journal of the Linnean Society*, 147(1), 91-98.
77. Marcussen, T., Heier, L., Brysting, A. K., Oxelman, B., & Jakobsen, K. S. (2015). From gene trees to a dated allopolyploid network: insights from the angiosperm genus *Viola* (Violaceae). *Systematic biology*, 64(1), 84-101.
78. Marcussen, T., Jakobsen, K. S., Danihelka, J., Ballard, H. E., Blaxland, K., Brysting, A. K., & Oxelman, B. (2012). Inferring species networks from gene trees in high-polyploid North American and Hawaiian violets (*Viola*, Violaceae). *Systematic biology*, 61(1), 107-126.
79. Mayers, A. M., & Lord, E. M. (1983). Comparative flower development in the cleistogamous species *Viola odorata*. I. A growth rate study. *American Journal of Botany*, 70(10), 1548-1555.
80. Mehrvarz, S. S., Vafi, M., & Marcussen, T. (2013). Taxonomic and anatomical notes on *Viola* sect. *Viola* (Violaceae) in Iran. *Wulfenia*, 20, 73-79.
81. Mered'a JR, P., Hodalova, I., Kučera, J., Zozomova-Lihova, J., Letz, D. R., & Slovak, M. (2011). Genetic and morphological variation in *Viola suavis* sl (Violaceae) in the western Balkan Peninsula: two endemic subspecies revealed. *Systematics and Biodiversity*, 9(3), 211-231.
82. Mered'a Jr, P., Hodálová, I., Mártonfi, P., Kučera, J., & Lihová, J. (2008). Intraspecific variation in *Viola suavis* in Europe: parallel evolution of white-flowered morphotypes. *Annals of Botany*, 102(3), 443-462.
83. Mereda, P., Jr., Hodalova, I., Kucera, J., Zozomova Lihova, J., Letz, D. R., & Slovak, M. (2011). *Viola suavis* isolate 247d ITS1 5.8S ITS2 sequence (Accession No. JF683833). GenBank, National Center for Biotechnology Information.
84. Metcalfe C.R. & Chalk L. (1950). *Anatomy of the Dicotyledons*. Vol. 1. – Oxford: Clarendon Press
85. Mohammadi Shahrestani, M., Saeidi Mehrvarz, S., Marcussen, T., & Yousefi, N. (2014). Taxonomy and comparative anatomical studies of *Viola* sect. *Sclerosium* (Violaceae) in Iran. *Acta botanica gallica*, 161(4), 343-353.

86. Moliner, C., Barros, L., Dias, M.I., Reigada, I., Ferreira, I.C., López, V., Langa, E. and Rincón, C.G. (2019). *Viola cornuta* and *Viola x wittrockiana*: Phenolic compounds, antioxidant and neuroprotective activities on *Caenorhabditis elegans*. *Journal of Food and Drug Analysis*, 27(4), 849-859.
87. Molyneux P. (2003). The use of the stable free radical diphenyl picrylhydrazyl (DPPH) for estimating antioxidant activity. *Songklanakarin J. Sci. Technol.*, 26(2):211-219
88. Muhammad, N., Saeed, M., Aleem, A., & Khan, H. (2012). Ethnomedicinal, phytochemical and pharmacological profile of genus *Viola*. *Phytopharmacology*, 3(1), 214-26.
89. Muriithi, A. N., Wamochi, L. S., & Njoroge, J. B. M. (2009). Effect of pH and magnesium on colour development and anthocyanin accumulation in tuberose florets. *African Crop Science Conference Proceedings*, Vol. 9. pp. 227-234.
90. Oprea, A. (2005). Lista critică a plantelor vasculare din România. Editura Universității „Alexandru Iona Cuza” Iași: 222 – 227
91. Padureanu, S. (2007). Characterization of morphology and germination capacity in *Viola odorata* L. pollen. *Analele Stiintifice ale Universitatii "Al. I. Cuza" din Iasi*, 53, 70.
92. Padureanu, S. (2007). Cytology of *Viola odorata* L. pollen germination. *Analele Stiintifice ale Universitatii "Al. I. Cuza" din Iasi*, 53, 60.
93. Pană, S., Cojuhari, T., Bacalov, I., Fărâmbă, V., & Topal, N. (2012). Compoziția chimică a plantelor medicinale din grădina botanică a muzeului național de etnografie și istorie naturală. Ghid teoretico-informativ de specialitate. *Buletin Științific. Revista de Etnografie, Științele Naturii și Muzeologie (Serie Nouă)*, 29(16), 104-127.
94. Parekh, J., Jadeja, D., & Chanda, S. (2005). Efficacy of aqueous and methanol extracts of some medicinal plants for potential antibacterial activity. *Turkish Journal of Biology*, 29(4), 203-210.
95. Pilberg, C., Ricco, M. V., & Alvarez, M. A. (2016). Foliar anatomy of *Viola maculata* growing in Parque Nacional Los Alerces, Chubut, Patagonia, Argentina. *Revista Brasileira de Farmacognosia*, 26, 459-463.
96. Pornon, A., Escaravage, N., Burrus, M., Holota, H., Khimoun, A., Andalo, C. (2016). *Viola canina* 18S rRNA gene and internal transcribed spacer 1 region (Accession No. KU974083). GenBank, National Center for Biotechnology Information.



97. Pr nting, M., L  v, C., Burman, R., G ransson, U. L. F., & Andersson, D. I. (2010). The cyclotide cycloviolacin O2 from *Viola odorata* has potent bactericidal activity against Gram-negative bacteria. *Journal of antimicrobial chemotherapy*, 65(9), 1964-1971.
98. Ramassamy, C. (2006). Emerging role of polyphenolic compounds in the treatment of neurodegenerative diseases: a review of their intracellular targets. *Eur J Pharmacol* 545:51–64
99. Rimkiene, S., Ragazinskiene, O., & Savickiene, N. (2003). The cumulation of Wild pansy (*Viola tricolor* L.) accessions: the possibility of species preservation and usage in medicine. *Medicina* (Kaunas), 39(4), 411-6.
100. Roberts, M. J. (2000). *Edible & medicinal flowers*. New Africa Books. New Africa Books: 58, 79
101. **Rosenhech, E.**, Lobiuc, A., & Zamfirache, M.-M. (2016). Phenolic contents and antioxidant activity in *Viola odorata* L., *V. tricolor* L. and *V. arvensis* (L.) Murray. *Analele Stiintifice ale Universitatii" Al. I. Cuza" din Iasi*, 62(1), 132-133.
102. **Rosenhech, E.**, Lobiuc, A., & Zamfirache, M.-M. - ( n curs de publicare) - The phytochemical potential of *Viola* L. species, *Viola* subgenus; *International Journal of Molecular Sciences*
103. **Rosenhech, E.**, Lobiuc, A., & Zamfirache, M.-M. ( n curs de publicare) - The phytochemical potential of *Viola* L. species, *Melanium* subgenus, subsect. *Bracteolatae*; *International Journal of Molecular Sciences*
104. **Rosenhech E.**, Lobiuc A., Boz I., Zamfirache M.-M., (2022) - Morpho-anatomical basis of discrimination between three *Viola* species; *Studia Universitatis „Vasile Goldiș” Seria Științele Vieții (Life Science Series)*, Vol. 32, issue 3.
105. Rubin, G., & Paolillo Jr, D. J. (1978). Vascular and general anatomy of the rootstocks of three stemless *Viola* species. *Annals of Botany*, 42(4), 981-988.
106. Saint-Lary, L., Roy, C., Paris, J. P., Tournayre, P., Berdagu , J. L., Thomas, O. P., & Fernandez, X. (2014). Volatile compounds of *Viola odorata* absolutes: identification of odorant active markers to distinguish plants originating from France and Egypt. *Chemistry & Biodiversity*, 11(6), 843-860.

- 107.Sârbu, I., Ștefan, N., & Oprea, A. (2013). Plante vasculare din România: determinant ilustrat de teren. Victor B victor. București: 452-460
- 108.Sârbu, I., Ștefan, N., Ivănescu, L., & Mânzu, C. (2001). Flora ilustrată a plantelor vasculare din estul României, II. Edit. Univ.“Al. I. Cuza” Iași. Book Review, 284-297.
- 109.Scoppola, A., & Lattanzi, E. (2012). Viola section Melanium (Violaceae) in Italy. New data on morphology of Viola tricolor-Group. Webbia, 67(1), 47-64.
- 110.Scoppola, A., Angeloni, D., & Franceschini, C. (2022). Classical morphometrics in V. arvensis and V. kitaibeliana (Viola sect. Melanium) reveals intraspecific variation with implications for species delimitation: Inferences from a case study in Central Italy. Plants, 11(3), 379.
- 111.Siatka, T., Kašparová, M. (2010). Seasonal variation in total phenolic and flavonoid contents and DPPH scavenging activity of Bellis perennis L. flowers. Molecules, 15:9450-9461
- 112.Simić, S., Aćimović, M., Vidović, S., Banožić, M., & Vladić, J. (2021). Viola odorata: Influence of supercritical fluid extraction on the efficiency of ultrasound-and microwave-assisted extraction of bioactive compounds. Croatian journal of food science and technology, 13(2), 191-200.
- 113.Słomka, A., Libik-Konieczny, M., Kuta, E., & Miszański, Z. (2008). Metalliferous and non-metalliferous populations of Viola tricolor represent similar mode of antioxidative response. Journal of plant physiology, 165(15), 1610-1619.
- 114.Słomka, A., Sutkowska, A., Szczepaniak, M., Malec, P., Mitka, J., & Kuta, E. (2011). Increased genetic diversity of Viola tricolor L.(Violaceae) in metal-polluted environments. Chemosphere, 83(4), 435-442.
- 115.Soare, L. C., Ferdeș, M., Dobrescu, C.-M. (2011). Species with potential for anthocyanins extraction in Argeș County flora. Muzeul Olteniei Craiova. Oltenia. Studii și comunicări. Științele Naturii. Tom. 27, No. 2
- 116.Tamura, K., Stecher, G., & Kumar, S. (2021). MEGA11: Molecular Evolutionary Genetics Analysis version 11. Molecular Biology and Evolution, 38(7), 3022–3027.  
<https://doi.org/10.1093/molbev/msab120>

117. Terashima, I., Miyazawa, S. I., & Hanba, Y. T. (2001). Why are sun leaves thicker than shade leaves? Consideration based on analyses of CO<sub>2</sub> diffusion in the leaf. *Journal of plant research*, 114, 93-105.
118. Terzano, R., Cuccovillo, G., Gattullo, C.E., Medici, L., Tomasi, N., Pinton, R., Mimmo, T. and Cesco, S. (2015). Combined effect of organic acids and flavonoids on the mobilization of major and trace elements from soil. *Biology and fertility of soils*, 51, 685-695.
119. Togor, G. C., & Burescu, P. (2013). Species-rich *Nardus* grasslands from the northern part of the Bihor Mountains. *Studia Universitatis" Vasile Goldis" Arad. Seria Stiintele Vietii (Life Sciences Series)*, 23(4), 505.
120. Toiu A, Oniga I, Tămaș M. (2009). Cercetări morfologice și anatomice asupra speciei *Viola tricolor* L. (Violaceae). *Revista medico-chirurgicală a Societății de Medici și Naturaliști din Iași*, 113 (2, supl. 4): 459-464.
121. Toiu, A., Oniga, I., & Tămaș, M. (2010). Morphological and anatomical researches on *Viola arvensis* Murray (Violaceae). *Farmacia*, 58(5), 654-659.
122. Toiu, A., Oniga, I., & Vlase, L. (2017). Determination of flavonoids from *Viola arvensis* and *V. declinata* (Violaceae). *Hop and Medicinal Plants*, 25(1/2), 125-130.
123. Toma, C. (2003). *Morfologia și anatomia plantelor. Manual de lucrări practice*. Ediția a 2-a revizuită și îmbunătățită, Edit. Universității Universității "Al. I. Cuza" din Iași.
124. Toma, C., & Ivanescu, L. (2013). *Plante vasculare din România. Determinator ilustrat de teren (Vascular plants of Romania. An illustrated field guide)*. *Analele Stiintifice ale Universității "Al. I. Cuza" din Iași*, 59(2), 109.
125. Tomescu, C. V., & Harasemciuc, T. I. (2018). Flora vasculară a rezervației "Codrul secular Loben"—județul Suceava. In *Horticultură, Viticultură și vinificație, Silvicultură și grădini publice, Protecția plantelor*, Vol. 47, pp. 427-434.
126. Tomović, G., Đurović, S., Buzurović, U., Niketić, M., Milanović, Đ., Mihailović, N., & Jakovljević, K. (2021). Accumulation of potentially toxic elements in *Viola* L. (Sect. *Melanium* Ging.) from the ultramafic and non-ultramafic soils of the Balkan Peninsula. *Water, Air, & Soil Pollution*, 232, 1-18.

127. Trusov, N. A. (2014). Aril morpho-anatomical structure and development of *Viola odorata* L. (Violaceae). *Modern Phytomorphology*, (6), 141-142.
128. Tutin, T.G., Heywood, V.H., Burges, N.A., Moore, D.M., Valentine, D.H., Walters, S.M., Webb, D.A. (1968). *Flora Europaea*, Vol 2 Rosaceae to Umbeliferae; At the University Press, Cambridge: 553-625
129. Vile, D., Gamier, E., Shipley, B., Laurent, G., Navas, M.L., Roumet, C., Lavorel, S., Diaz, S., Hodgson, J.G., Lloret, F., & Midgley, G.F. (2005). Specific leaf area and dry matter content estimate thickness in laminar leaves. *Annals of botany*, 96(6), 1129-1136.
130. Vukics, V., Hevesi Toth, B., Ringer, T., Ludanyi, K., Kery, A., Bonn, G. K., & Guttman, A. (2008b). Quantitative and qualitative investigation of the main flavonoids in heartsease (*Viola tricolor* L.). *Journal of chromatographic science*, 46(2), 97-101.
131. Vukics, V., Kery, A., Bonn, G. K., & Guttman, A. (2008a). Major flavonoid components of heartsease (*Viola tricolor* L.) and their antioxidant activities. *Analytical and bioanalytical chemistry*, 390, 1917-1925.
132. Watson, J. M., Flores, A. R., Nicola, M. V., & Marcussen, T. (2021). *Viola* subgenus *Andinium*, preliminary monograph. Scottish Rock Garden Club with International Rock Gardener, 1-215.
133. Watson, L., Dallwitz, M.J. (1992). The families of flowering plants: descriptions, illustrations, identification, and information retrieval; Version: 19th October 2013; <http://delta-intkey.com>. accesat în martie 2018
134. Wellburn, A. R. (1994). The spectral determination of chlorophylls a and b, as well as total carotenoids, using various solvents with spectrophotometers of different resolution. *Journal of plant physiology*, 144(3), 307-313.
135. Wiklund, C. (1984). Egg-laying patterns in butterflies in relation to their phenology and the visual apparency and abundance of their host plants. *Oecologia*, 23-29.
136. Winkel-Shirley, B. (2002). Biosynthesis of flavonoids and effects of stress. *Current opinion in plant biology*, 5(3), 218-223.
137. Witkowska-Banaszczak, E., Bylka, W., Maławska, I., Goślińska, O., & Muszyński, Z. (2005). Antimicrobial activity of *Viola tricolor* herb. *Fitoterapia*, 76(5), 458-461.

138. Wrolstad, R. E., Durst, R. W., & Lee, J. (2005). Tracking color and pigment changes in anthocyanin products. *Trends in Food Science & Technology*, 16(9), 423-428.
139. Yockteng, R., Jr Ballard, H. E., Mansion, G., Dajoz, I., & Nadot, S. (2003). Relationships among pansies (*Viola* section *Melanium*) investigated using ITS and ISSR markers. *Plant Systematics and Evolution*, 241, 153-170.
140. Yousefi, N., Mehrvarz, S. S., & Marcussen, T. (2012). Anatomical studies on selected species of *Viola* (Violaceae). *Nordic Journal of Botany*, 30(4), 461-469.
141. Yousefi, N., Sajedi, R. H., Marcussen, T., Saeidi Mehrvarz, S. (2010). *Viola kitaibeliana* ITS1 5.8S rRNA partial sequence (Accession No. HM756253). GenBank, National Center for Biotechnology Information.
142. Zamfirache MM. (2005). Fiziologie vegetală. 1, Edit. Azimuth, Iași, p. 20-21, 72-83
143. Zamfirache, MM., Stratu, A., Olteanu, Z., Galeș, R. (1997). Fiziologie vegetală – Ghid de lucrări practice”. Edit. Universității Alexandru Ioan Cuza”, Iași, Cap. 7, pag. 4-5
144. Zarrabi, M., Dalirfardouei, R., Sepehrizade, Z., & Kermanshahi, R. K. (2013). Comparison of the antimicrobial effects of semipurified cyclotides from Iranian *Viola odorata* against some of plant and human pathogenic bacteria. *Journal of applied microbiology*, 115(2), 367-375.
145. Zhang, J., Wang, L. S., Gao, J. M., Xu, Y. J., Li, L. F., & Li, C. H. (2012). Rapid separation and identification of anthocyanins from flowers of *Viola yedoensis* and *V. prionantha* by high-performance liquid chromatography–photodiode array detection–electrospray ionisation mass spectrometry. *Phytochemical Analysis*, 23(1), 16-22.
146. Zhou, Y., Zheng, J., Li, Y., Xu, D. P., Li, S., Chen, Y. M., & Li, H. B. (2016). Natural polyphenols for prevention and treatment of cancer. *Nutrients*, 8(8), 515.
147. Zimniewska, M., Pawlaczyk, M., Krucinska, I., Frydrych, I., Mikolajczak, P., Schmidt-Przewozna, K., Komisarzyk, A., Herczynska, L., Romanowska, B. (2019). The influence of natural functional clothing on some biophysical parameters of the skin. *Textile Research Journal*, 89(8), 1381-1393.